REFERENCE NUMBER:_____

VETERINARY HEALTH CERTIFICATE
EXPORT OF BOVINE EMBRYOS TO NEW ZEALAND

I.1 Consignor (Exporter): Name: Address:	I.2 Certificate reference number: I.3. Competent Authority: **CANADIAN FOOD INSPECTION AGENCY**
I.4. Consignee (Importer): Name: Address:	
I.5. Country of origin: **CANADA** ISO Code: **CA**	I.6 Zone or compartment of origin**
I.7. Country of destination: **NEW ZEALAND** ISO code: **NZ**	I.8 Zone or compartment of destination**
I.9. Place of origin: Name: Address:	
I.10. Place of shipment:	I.11. Date of departure:
I.12. Means of transport ☐ Aeroplane ☐ Ship Identification:	I.13. Expected border post: I.14. CITES permit No(s): N/A
I.15. Description of commodity: **FROZEN BOVINE EMBRYOS**	I.16. Commodity Code (HS Code): **0511 9991** I.17. Total number of embryos:
I.18. Temperature of commodities for transport: -196 ºC	I.19. Total number of packages:
I.20. Identification of container(s) and seal number: Container _____ Seal Number _____ _____ _____ _____ _____	I.21. Type of packaging:

** If referenced in Table 1

HEALTH REQUIREMENTS

I, the undersigned Official Veterinarian, duly authorised by the Government of Canada, hereby certify, after due enquiry and to the best of my knowledge and belief, that:

1. Eligibility

1.1 Frozen, in vivo derived embryos from the Bovinae subfamily

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2. Diagnostic tests, vaccines and treatment: (*Strikeout unused options and initial)

2.1 All pre-export and/or surveillance testing required by this veterinary certificate has been:

2.1.1 *conducted by a laboratory approved by the Competent Authority of the exporting country; Or

2.1.2 *conducted by a laboratory approved by the Competent Authority of any other country approved to export the specified type of germplasm to New Zealand.

2.2 All laboratory samples required by this veterinary certificate have been collected, processed, and stored in accordance with the OIE's recommendations or as described in Approved Diagnostic Tests, Vaccines, Treatments and Post-Arrival Testing Laboratories for Animal Import Health Standards, MPI-STD-TVTL(1).

2.3 All diagnostic test(s) and vaccines that are required below are those that have been approved by Ministry for Primary Industries (MPI) for that purpose and documented in MPI-STD-TVTL.

2.4 All products and vaccinations required by this veterinary certificate to be administered to meet the specific disease requirements below have been administered according to the manufacturer's instruction.

2.5 Any vaccine(s) administered to satisfy import requirements was/were either the final dose of a primary vaccination course has been administered or the recommended booster to complement the primary course.

3. Donor requirements

3.1 The embryo donors met the recommendations in the *Code chapter Collection and Processing of In Vivo Derived Embryos from Livestock and Equids*, and the other specified requirements in this veterinary certificate.

3.2 The embryo donors were resident in the embryo collection herd for at least 28 days prior to embryo collection for export to New Zealand. While resident with the collection herd, the herd was not subject to veterinary restrictions for organisms managed in this veterinary certificate.

3.3 Embryo donors were legally imported a minimum of 60 days prior to embryo collection, and lived only in countries approved to export bovine germplasm to New Zealand. (USA, EU, United Kingdom, Australia, Switzerland)

3.4 On the day of embryo collection, the veterinary member of the approved embryo collection team determined that the donors were free from clinical evidence of infectious diseases transmissible in germplasm.

3.5 Where a specific requirement of this veterinary certificate was met by pre-collection testing, the embryo donors were isolated from other animals not of equivalent tested health status, from the time of the test sample collection until completion of embryo collection for export.

3.6 Where a specific requirement of this veterinary certificate for a risk organism was met by monitoring the embryo donors for clinical signs for a specified time post collection, the embryos were stored for that amount of time prior to export.

4. Embryo collection, processing, and storage (*Strikeout unused options and initial)

4.1 Embryos were collected, washed, processed, stored and traceability maintained under the supervision of an embryo collection team veterinarian:

4.1.1 approved by the Canadian Food Inspection Agency, under the Embryo Export Approval Program

4.1.2 in accordance with the recommendations in the OIE Code chapter on *Collection and Processing of In Vivo Derived Embryos from Livestock and Equids (2015)*;

4.1.3 in accordance with the recommendations in the IETS Manual.

4.2 The embryo collection team operated in accordance with the conditions listed in the OIE Code chapter on *Collection and Processing of In Vivo Derived Embryos from Livestock and Equids (2015)*.

4.3 At the time of embryo collection each embryo was examined over its entire surface at not less than 50X magnification and found to have an intact zona pellucida and be free of adherent material.

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4.4 *If any embryo in the consignment underwent micro-manipulation that caused a breach of the zona pellucida it was performed as per the procedures described in the OIE Code chapter *Collection and Processing of Micromanipulated Oocytes or Embryos from Livestock and Horses (2009)* and the IETS Manual.

4.5 All biological products of animal origin used in the media and solutions for collection, processing, washing or storage of the embryos were free of pathogenic organisms including pestiviruses and prions. Media and solutions were sterilised by approved methods according to the IETS Manual and handled in a manner that ensured that sterility is maintained.

4.6 All straws were sealed, and clearly and permanently marked to identify the donor and the date(s) of freezing. The markings conform to international standards of the International Committee for Animal Recording (ICAR) and the IETS Manual.
*If a code is used for this information, its decipher instructions accompany the consignment.

4.7 The embryos have been stored and transported only with germplasm that has been collected and processed in accordance with the OIE Code.

4.8 The embryos have been stored at all times at a location approved by the Competent Authority of the exporting country until the time of export.

4.9 Subject to 4.10, the embryos are exported directly from the country in which they were collected or transited through a country that has an agreed veterinary certificate for bovine embryo export to New Zealand.

4.10 *If embryos were collected in a country that has an agreed veterinary certificate for bovine embryo export to New Zealand and stored in another country that also has an agreed veterinary certificate for bovine embryos to New Zealand (country of storage), the embryos are accompanied by:

4.10.1 A declaration from the Competent Authority of the country of storage that:

4.10.1.1 identifies the embryos from the country of origin as the embryos being exported to New Zealand;

4.10.1.2 certifies that the embryos have been stored and transported in the country of storage in accordance with the requirements of this veterinary certificate;

4.10.2 Evidence that the embryos meet the rest of the import requirements in the form of either:

4.10.2.1 *A veterinary certificate issued by the Competent Authority of the origin country; or

4.10.2.2 *A letter from the Competent Authority of the origin country confirming the embryos meet the requirements of their agreed veterinary certificate and indicating which requirements therein have been fulfilled.

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5. Transport (*Strikeout unused options and initial)

5.1 The transport container in which the embryos are to be transported to New Zealand is:

5.1.1 new*; or

5.1.2 disinfected and is free of contamination*;
5.1.2.1 2% available chlorine (e.g. chlorine bleach)*;
5.1.2.2 2% Virkon*;
5.1.2.3 2.4% Prevail (dilution ratio 1:40)*;

Date of disinfection (yyyyy/mm/dd)___

5.2 All transport containers in which embryos are transported to New Zealand have been sealed, by an Official Veterinarian, using tamper-evident seals that are positioned to ensure that no germplasm can be added after the transport container has been sealed.

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SPECIFIED REQUIREMENTS

6 Bovine viral diarrhoea virus genotype 2 (BVDV2)

6.1 A sample of the unfiltered collection fluid or an embryo from the collection for export to New Zealand was tested for BVDV2 by virus isolation with negative results; or

6.2 The embryo donor was tested for persistent BVDV2 infection (Virus Isolation or Antigen capture ELISA test), with negative results; and the

6.2.1 The semen used to produce the embryo satisfies the BVDV2 requirements of the OIE Animal Health Code Chapter on Collection and Processing of Bovine, Small Ruminant and Porcine Semen, 2021 version.

6.2.2 The embryo donor was not vaccinated against BVDV2 in the last 30 days;

6.2.3 The embryo donor was tested for acute BVDV2 infection, with negative results, in one of the following ways:

6.2.3.1 with antigen capture ELISA immediately prior to an isolation period of at least 21 days before collection for New Zealand. Isolation must exclude cattle that were not tested negative for BVDV2 upon entry to the collection herd, and throughout isolation the herd showed no clinical signs consistent with BVDV2; or

6.2.3.2 with virus isolation within 48 hours before or after embryo collection for New Zealand; or

6.2.3.3 serologically with an antibody test between 2 weeks and 6 months after embryo collection.

7. Foot and Mouth Disease (FMD)

7.1 The donor dam was resident for at least the 3 months before embryo collection in a country or zone that was free from FMD without vaccination in accordance with the OIE Code.

8. Lumpy Skin Disease (LSD)

8.1 The donor dam was resident for 6 months prior to germplasm collection in a country or zone that was free of LSD as defined by the OIE Code.

9. Rift Valley Fever (RVF)

9.1 The donor dam was resident, for at least the 30 days prior to, and during embryo collection for export to New Zealand in a country or zone that was free from RVF in accordance with the OIE Code.

10. *Coxiella burnetii* (Q-fever)

10.1 The donor dam has never been confirmed positive for Q fever; and

10.1.1 was subjected to a serological test for Q fever (ELISA) as described in MPI-STD-TVTL, on a sample collected between 21 and 120 days after each embryo collection for export to New Zealand, with negative results.

11. *Leptospira interrogans serovar harjoprajitno* (leptospirosis) (*Strikeout unused options and initial)

11.1 *Antibiotics were added to either collection or all washing media using one of the combinations below; or

11.1.1 50 IU/ml penicillin and 50 µg/ml streptomycin*; or
11.1.2 50 µg/ml tylosin*.

11.2 *The donor dam was:

11.2.1 given a single injection of oxytetracycline (20 mg/kg of body weight IM) between 2 and 10 days prior to embryo collection for New Zealand.

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12. Mycobacterium tuberculosis (bovine tuberculosis)

12.1 No clinical signs of bovine tuberculosis were observed in the embryo collection herd during the 24 hours prior to embryo collection for export to New Zealand;

12.2 The donor was from an embryo collection herd that was free at the time of collection for export to New Zealand from bovine tuberculosis in accordance with the Competent Authority of the exporting country; and either

12.2.1 from a country or zone free from bovine tuberculosis.

13. Mycoplasma mycoides subspecies mycoides SC (contagious bovine pleuropneumonia, CBPP)

13.1 The germplasm donor was born in and had been continuously resident in a country free from CBPP

14. *Mycoplasma bovis* (*Strikeout unused options and initial)

14.1 Collection and processing of embryos was in accordance with the recommendations of the OIE Code, with the modifications indicated below:

14.1.1 The embryos were subjected to the treatment protocol described in the IETS Manual; 5th edition, Volume 1 Chapter 6 – Antibiotic Usage in Embryo Production. Embryos were incubated at 37°C in media containing tylosin (200 µg/mL) for a minimum of 4 hours after being washed 10 times; or

14.2 Each embryo collection for export to New Zealand was tested with a validated PCR test for *M. bovis* as described in MPI-STD-TVTL, with negative results

Sample

_____ _____ _____

Name of Approved Team Veterinarian | ET Code | Signature of Approved Team Veterinarian

_____ _____

Date (yyyy-mm-dd) | Signature of Official Veterinarian
Canadian Food Inspection Agency

_____

Name of Official Veterinarian (in block letters)

Address:_____

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Note: Official Veterinarian signature and Official Stamp must be applied to each page

REFERENCE NUMBER:_____

Table I: Embryo Information (date format is yyyy-mm-dd)

Breed	Female Donor registration number	Donor sire registration number	Date of entry into Embryo Collection Herd/Centre (yyyy-mm-dd)	Date of collection (yyyy-mm-dd)	Freezing date (yyyy-mm-dd)	Number of straws	Name and Address of Embryo Collection Herd/Centre

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REFERENCE NUMBER:_____

Table II: Test Information (date format is yyyy-mm-dd.)

Female Donor Registration number	Date of collection (yyyy-mm-dd)	Bovine tuberculosis intradermal tuberculin test	Bovine viral diarrhoea (BVD)			Q fever			Leptospirosis			*Mycoplasma bovis*		
			Sampling date	Test type	Result	Sampling date	Test type	Result	Sampling date	Test type	Result	Sampling date	Test type	Result

Sample

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